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## Original Article

# Synthesis, radiolabelling and *in vitro* and *in vivo* evaluation of a novel fluorinated ABP688 derivative for the PET imaging of metabotropic glutamate receptor subtype 5

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**Abstract:** (*E*)-3-(Pyridin-2-ylethynyl)cyclohex-2-enone *O*-(2-(3-<sup>18</sup>F-fluoropropoxy)ethyl) oxime ([<sup>18</sup>F]-PSS223) was evaluated *in vitro* and *in vivo* to establish its potential as a PET tracer for imaging metabotropic glutamate receptor subtype 5 (mGluR5). [<sup>18</sup>F]-PSS223 was obtained in 20% decay corrected radiochemical yield whereas the non-radioactive PSS223 was accomplished in 70% chemical yield in a S<sub>N</sub>2 reaction of common intermediate mesylate 8 with potassium fluoride. The *in vitro* binding affinity of [<sup>18</sup>F]-PSS223 was measured directly in a Scatchard assay to give K<sub>d</sub> = 3.34 ± 2.05 nM. [<sup>18</sup>F]-PSS223 was stable in PBS and rat plasma but was significantly metabolized by rat liver microsomal enzymes, but to a lesser extent by human liver microsomes. Within 60 min, 90% and 20% of [<sup>18</sup>F]-PSS223 was metabolized by rat and human microsome enzymes, respectively. *In vitro* autoradiography on horizontal rat brain slices showed heterogeneous distribution of [<sup>18</sup>F]-PSS223 with the highest accumulation in brain regions where mGluR5 is highly expressed (hippocampus, striatum and cortex). Autoradiography *in vitro* under blockade conditions with ABP688 confirmed the high specificity of [<sup>18</sup>F]-PSS223 for mGluR5. Under the same blocking conditions but using the mGluR1 antagonist, JNJ16259685, no blockade was observed demonstrating the selectivity of [<sup>18</sup>F]-PSS223 for mGluR5 over mGluR1. Despite favourable *in vitro* properties of [<sup>18</sup>F]-PSS223, a clear-cut visualization of mGluR5-rich brain regions *in vivo* in rats was not possible mainly due to a fast clearance from the brain and low metabolic stability of [<sup>18</sup>F]-PSS223.

**Keywords:** mGluR5, PET imaging, [<sup>18</sup>F]-PSS223, [<sup>11</sup>C]-ABP688, [<sup>18</sup>F]-FDEGPECO, autoradiography, microsome enzymes

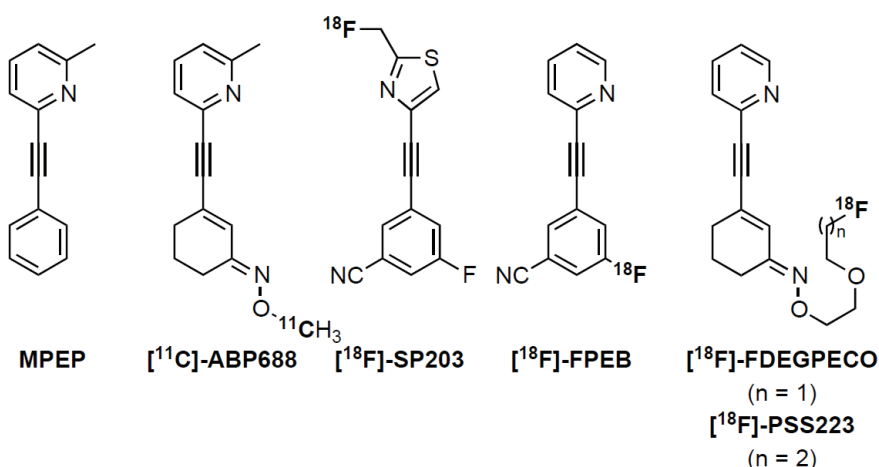
## Introduction

Positron emission tomography (PET) is a powerful non-invasive imaging technique which allows quantification of biochemical and pharmacodynamic processes in healthy and diseased states. In drug discovery and development, PET offers opportunities to investigate drug-target interactions *in vivo* and to monitor the effects of a drug candidate on the progression of a disease [1, 2]. The design of PET probes which selectively target a particular receptor, enzyme or transporter is a major challenge in further advancement of PET technique. Besides analogues of endogenous substrates such as [<sup>18</sup>F]-

fluorodeoxyglucose ([<sup>18</sup>F]-FDG) or [<sup>11</sup>C]- or [<sup>18</sup>F]-labelled amino acids [3, 4], small synthetic organic molecules are emerging as promising scaffolds for the development of highly selective PET probes.

Metabotropic glutamate receptor subtype 5 (mGluR5) is a seven transmembrane domain, G-protein coupled receptor, belonging to group I of the metabotropic glutamate receptors and it is mainly located postsynaptically [5-7]. mGluR5 is involved in long-term potentiation processes [8] and plays a role in several disorders of central nervous system (CNS) such as schizophrenia, depression, anxiety, Alzheimer and Parkinson's

## Development of novel mGluR5 PET tracer



**Figure 1.** Structures of MPEP and selected PET tracers for imaging of mGluR5.

disease [9-14]. It is considered an important future drug target and much attention has been given to the development of an optimal PET tracer for imaging of mGluR5 [15].

It was not until the development of synthetic mGluR5 antagonist 2-methyl-6-(phenylethynyl)pyridine (MPEP) [16, 17], which unlike amino acid derived ligands acts via the non-competitive mechanism and not at the conserved glutamate binding site, that significant research efforts were made. This led to the development of (*E*)-3-((6-methylpyridin-2-yl)ethynyl)cyclohex-2-en-1-one- $O$ - $^{11}\text{C}$ -methyl oxime ( $[^{11}\text{C}]\text{-ABP688}$ , **Figure 1**) by the Ametamey group [18, 19] which is to date the most successful clinically applied mGluR5 PET tracer. Despite its excellent properties as a PET imaging agent in human subjects [19],  $[^{11}\text{C}]\text{-ABP688}$  does have one limitation and this is due to the short physical half-life (20 min) of carbon-11 which limits its application to facilities with an on-site cyclotron. This has provoked further research efforts towards the development of  $^{18}\text{F}$ -labelled PET tracers, which have a longer physical half-life (109.8 min). To date, the two most successful mGluR5  $^{18}\text{F}$ -labelled PET tracers are 3-fluoro-5-(2-(2- $[^{18}\text{F}]\text{-(fluoromethyl)-thiazol-4-yl)-ethynyl)benzonitrile$  ( $[^{18}\text{F}]\text{-SP203}$ ) developed by the Pike group [20-22] and 3- $[^{18}\text{F}]\text{fluoro-5-(2-pyridinylethynyl)benzonitrile}$  ( $[^{18}\text{F}]\text{-FPEB}$ ) from the Hamill group [23-25] (**Figure 1**). However, the first undergoes significant defluorination in rodents and monkeys and to a lesser extent in humans and the latter is typically obtained in

low radiochemical yields.

Our group aimed to develop a fluorine-18 analogue of  $[^{11}\text{C}]\text{-ABP688}$  with the same excellent imaging properties as  $[^{11}\text{C}]\text{-ABP688}$ , but with added advantage of a longer physical half-life. Several derivatives were prepared [26, 27] and evaluated which finally led to the development of (*E*)-3-(pyridin-2-ylethynyl)cyclohex-2-en-1-one  $O$ -(2-(2- $^{18}\text{F}$ -fluoroethoxy)ethyl) oxime ( $[^{18}\text{F}]\text{-FDEGPeco}$ ) [28, 29], which maintained the same  $[^{11}\text{C}]\text{-ABP688}$  scaffold, but the oxime functionality was additionally adorned with a six-atom long lipophilic chain (as oppose to a methyl in  $[^{11}\text{C}]\text{-ABP688}$ ) [1, 2, 30, 31]. The *in vivo* evaluation of  $[^{18}\text{F}]\text{-FDEGPeco}$  allowed visualization of mGluR5 in the rat brain, albeit with a lower relative accumulation and shorter residence time in mGluR5-rich brain regions than observed for  $[^{11}\text{C}]\text{-ABP688}$ , which prompted us to re-evaluate the structure of  $[^{18}\text{F}]\text{-FDEGPeco}$ . The lipophilicity of  $[^{18}\text{F}]\text{-FDEGPeco}$  is significantly lower than that of  $[^{11}\text{C}]\text{-ABP688}$  [18], and we chose to pursue an  $[^{18}\text{F}]\text{-FDEGPeco}$  analogue (*E*)-3-(Pyridin-2-ylethynyl)cyclohex-2-en-1-one  $O$ -(2-(3- $^{18}\text{F}$ -fluoropropoxy)ethyl) oxime ( $[^{18}\text{F}]\text{-PSS223}$ , **Figure 1**) in which the side chain functionality is extended by one methylene group with the aim to increase the lipophilicity towards that of  $[^{11}\text{C}]\text{-ABP688}$  [28, 32].

In this manuscript we report on the synthesis, radiolabelling, *in vitro* and *in vivo* evaluation of this novel compound ( $[^{18}\text{F}]\text{-PSS223}$ ). The PET images of ( $[^{18}\text{F}]\text{-PSS223}$ ) are also compared with

those obtained with [ $^{11}\text{C}$ ]-ABP688 and [ $^{18}\text{F}$ ]-FDEGPECO.

## Materials and methods

### General

All reactions requiring anhydrous conditions were conducted in flame-dried glass apparatus under an atmosphere of inert gas. All chemicals and anhydrous solvents were purchased from Aldrich or ABCR and used as received unless otherwise noted. [ $^3\text{H}$ ]-ABP688 (2.405 GBq/mmol, 37 MBq/mL solution in EtOH) was obtained from AstraZeneca. 3-Ethynylcyclohex-2-enone, (*E*)-3-ethynylcyclohex-2-enone oxime and (*E*)-3-(pyridin-2-ylethynyl)cyclohex-2-enone oxime (**5**) were prepared as previously reported and the characterization data is in complete agreement with those previously reported [28, 33]. To obtain (*E*)-3-((6-Methylpyridin-2-yl)ethynyl)cyclohex-2-enone O- $^{11}\text{C}$ -methyl oxime ([ $^{11}\text{C}$ ]-ABP688) radiolabelling was performed using module system as previously described [18, 34].

Preparative chromatographic separations were performed on Aldrich Science silica gel 60 (35–75  $\mu\text{m}$ ) and reactions followed by TLC analysis using Sigma-Aldrich silica gel 60 plates (2–25  $\mu\text{m}$ ) with fluorescent indicator (254 nm) and visualized with UV or potassium permanganate.

Infrared spectra were recorded on a JASCO FT/IR 6200 (OmniLab) spectrometer using a chloroform solution of compound.  $^1\text{H}$  and  $^{13}\text{C}$  NMR spectra were recorded in Fourier transform mode at the field strength specified on Bruker Avance FT-NMR spectrometers. Spectra were obtained from the specified deuterated solvents in 5 mm diameter tubes. Chemical shift in ppm is quoted relative to residual solvent signals calibrated as follows:  $\text{CDCl}_3$   $\delta_{\text{H}}$  ( $\text{CHCl}_3$ ) = 7.26 ppm,  $\delta_{\text{C}}$  = 77.2 ppm;  $(\text{CD}_3)_2\text{SO}$   $\delta_{\text{H}}$  ( $\text{CD}_3\text{SOCHD}_2$ ) = 2.50 ppm,  $\delta_{\text{C}}$  = 39.5 ppm. Multiplicities in the  $^1\text{H}$  NMR spectra are described as: s = singlet, d = doublet, t = triplet, q = quartet, quint. = quintet, m = multiplet, b = broad; coupling constants are reported in Hz. Numbers in parentheses following carbon atom chemical shifts refer to the number of attached hydrogen atoms as revealed by the DEPT spectral editing technique. Electrospray (ES) mass spectra (LRMS) were obtained with a Micromass Quattro micro API LC electrospray ionization and electrospray (ES)

mass spectra (HRMS) were obtained with a Bruker FTMS 4.7 T BioAPEXII spectrometer. Electron-impact (EI) and chemical ionisation (CI) mass spectra (LRMS and HRMS) were obtained with a Waters Micromass AutoSpec Ultima MassLynx 4.0 spectrometer. Ion mass/charge ( $m/z$ ) ratios are reported as values in atomic mass units.

Semi-preparative purification of radiolabelled material was performed on a Merck-Hitachi L6200A system equipped with Knauer variable wavelength detector and an Eberline radiation detector using a reverse phase column (C18 Phenomenex Gemini, 5 mm, 250x10 mm) and eluting with gradient: 0–5 min 5% aq. MeCN, 5–15 min 5–50% aq. MeCN, 15–30 min 50% aq. MeCN, 30–50 min, 50–90% aq. MeCN, 50–65 min, 65% MeCN at flow rate 5 mL/min. Analytical HPLC samples were analyzed by Agilent HPLC 1100 system equipped with UV multi-wavelength detector and Raytest Gabi star radiation detector using reverse phase column (ACE 111-0546, C18, 3 mm, 50x4.6 mm) and eluting with 45% aq. MeCN at flow rate 1 mL/min. Samples for PBS, plasma and microsome stability were analyzed by Waters ultra-performance liquid chromatography (UPLC<sup>TM</sup>) system equipped with Berthold coincidence detector (FlowStar LB513) using UPLC column (Waters Acquity BEH C18, 1.7 mm, 50x2.1 mm) and eluting with gradient 0–70% aq. MeCN over 5 min at flow 0.7 mL/min or for microsome assay with 30% aq. MeCN over 5 min at flow 0.7 mL/min.

Bovine serum albumin was purchased from Acros Organics. Pooled human liver microsomes, pooled liver microsomes from male Spargue Dawley rats (20 mg protein per mL) and NADPH regenerating system (A: 31 mM  $\text{NADP}^+$ , 66 mM glucose-6-phosphate, 66 mM  $\text{MgCl}_2$ ; B: 40 U/mL glucose-6-phosphate dehydrogenase) were obtained from BD Biosciences.

Male Wistar rats were obtained from Charles River (Sulzfeld, Germany). Animal care and all experimental procedures were approved by the Cantonal Veterinary Office in Zurich, Switzerland. The animals were allowed free access to food and water.

### Radiosynthesis

(*E*)-3-(Pyridin-2-ylethynyl)cyclohex-2-enone O-(2-

(3-<sup>18</sup>F-fluoropropoxy)ethyl oxime ([<sup>18</sup>F]-PSS223): No-carrier-added [<sup>18</sup>F]-fluoride was produced via nuclear <sup>18</sup>O(p, n)<sup>18</sup>F reaction from enriched <sup>18</sup>O-water using IBA cyclone 18/9 cyclotron and it was immediately trapped on a QMA cartridge (preconditioned with 0.5 M aq. K<sub>2</sub>CO<sub>3</sub> (1x5 mL), then H<sub>2</sub>O (1x5 mL) and dried in air). The trapped [<sup>18</sup>F]-fluoride was eluted from the cartridge with 0.25% wt Kryptofix-222® solution (1 mL) in basic (0.05% wt K<sub>2</sub>CO<sub>3</sub>) aq MeCN (75% v/v) into a tightly closed reaction vial. The solvents were evaporated *in vacuo* (130 mbar) with gentle stream of N<sub>2</sub> gas at 110 °C over 5 min. To the resulting solid residue anhydrous MeCN (1 mL) was then added and the mixture was azeotropically dried *in vacuo* (130 mbar) with gentle stream of N<sub>2</sub> at 110 °C. To the dried Kryptofix-222®/[<sup>18</sup>F]- complex a solution of (E)-3-(2-(((3-(pyridin-2-ylethynyl)cyclohex-2-en-1-ylidene)amino)oxy)ethoxy)propyl methanesulfonate (2.44 mg, 6.22 μmol) in anhydrous N,N'-dimethylformamide (0.3 mL) was added and the dark brown mixture was heated at 90 °C for 10 min. The crude mixture was diluted with 50% v/v aq. MeCN (2 mL) and purified via semi-preparative HPLC. The desired product was collected (retention time: 31.9 min) and immediately diluted with H<sub>2</sub>O (10 mL). The aqueous solution was passed through a C18 cartridge (preconditioned with EtOH (1x5 mL), then H<sub>2</sub>O (1x5 mL) and dried in air) and the cartridge was washed with H<sub>2</sub>O (2x1 mL) and the product was eluted from the C18 cartridge with EtOH (1x0.3 mL) in a sterile vial containing 50% aq. PEG200 (5 mL) to afford the radiolabelled title compound in 20% decay corrected yield. Typically starting from ca. 30 GBq of activity, 5-6 GBq of product was obtained. The radiochemical purity was >97% and specific activity in the range of 100-320 GBq/mmol.

#### Determination of logD

For purposes of logD determination a further product elution from the C18 cartridge was carried out using EtOH (0.3 mL) and the solvent was evaporated *in vacuo* (100 mbar) at 90 °C for 5 min.

The determination of logD value was performed using the shake-flask method as previously reported [28, 35]. [<sup>18</sup>F]-PSS223 (453 MBq) was partitioned between phosphate buffer (pH 7.4) saturated with 1-octanol (2 mL) and 1-octanol saturated with phosphate (pH 7.4) buffer (2

mL). The octanol (top) phase (0.5 mL) was washed with phosphate (pH 7.4) buffer saturated with 1-octanol (2x0.5 mL). The purity of the material (14 MBq) was confirmed by HPLC analysis. Finally washed octanol phase (0.1 mL) was diluted with phosphate (pH 7.4) buffer saturated with 1-octanol (0.1 mL) and the two phases were shaken and radioactivity in each phase was measured in a γ-counter.

#### Pharmacology

##### Competition Binding Assay

Brain membranes were prepared from Sprague Dawley rat brains as described previously [28]. Frozen membranes were thawed on ice and pelleted at 45000xg at 4 °C for 5 min. The membranes were washed twice with HEPES buffer (30 mM HEPES, 110 mM NaCl, 5 mM KCl, 2.5 mM CaCl<sub>2</sub>, 1.2 mM MgCl<sub>2</sub>, pH 8 at 4 °C) and resuspended in HEPES buffer at a protein concentration of 1.3 mg/mL. The binding assay was performed as previously described [28]. In brief, brain membranes (0.1 mg protein) were incubated in triplicates at ambient temperature with 2 nM [<sup>3</sup>H]ABP688 and PSS223 at concentrations between 10 pM and 100 μM in a total volume of 0.2 ml HEPES. PSS223 was diluted from a 1 mM ethanolic (50%) solution. The corresponding EtOH concentrations did not affect [<sup>3</sup>H]ABP688 binding (data not shown). Unspecific binding of [<sup>3</sup>H]ABP688 was estimated with 100 μM MMPEP. After 45 min, the samples were filtrated and the filters containing the membranes with bound [<sup>3</sup>H]ABP688 were measured in a β-counter (Beckman LS6500). Bound [<sup>3</sup>H]ABP688 (B, pmol per mg protein) was fitted with Excel solver to **Equation 1** to estimate IC<sub>50</sub>.

$$B = B_{\min} + ((B_{\max} - B_{\min}) / (1 + (C / IC_{50}))) \quad \text{Eq. 1}$$

where C is the total PSS223 concentration, B<sub>max</sub> is the maximal B, i.e., the plateau in the B/C plot at low log C and B<sub>min</sub> is the minimal B, i.e., the plateau at high log C. The inhibition constant K<sub>i</sub> of PSS223 was estimated from IC<sub>50</sub> and K<sub>d</sub> of ABP688 (1.7±0.2 nM) [18] with the Cheng-Prusoff equation.

##### Saturation Binding Assay

Brain membranes were incubated with [<sup>18</sup>F]-PSS223 at concentrations between 0.25 and

100 nM as described above. Unspecific binding was determined with 100  $\mu$ M MMPEP. After filtration, bound [ $^{18}$ F]-PSS223 was quantified in a  $\gamma$ -counter (PerkinElmer, Wizard) and  $K_d$  was estimated according to **Equation 2**,

$$B_{\text{spec}} = (B_{\text{max,spec}} \times 1/K_d \times C_u) / (1 + 1/K_d \times C_u) \quad \text{Eq. 2}$$

where  $B_{\text{spec}}$  is the difference between B and the unspecific binding (at 100  $\mu$ M MMPEP) and  $C_u$  is the unbound concentration, i.e. the difference between the total and the bound concentrations.  $B_{\text{max,spec}}$  is the maximal  $B_{\text{spec}}$  at receptor saturation. Three independent experiments were performed with [ $^{18}$ F]-PSS223 obtained from three independent radiolabelling productions.

#### *In vitro autoradiography*

Frozen horizontal brain slices (20 mm) from a male Wistar rat (492 g) adsorbed to SuperFrost Plus slides were thawed at ambient temperature and preincubated on ice for 10 min in HEPES buffer (see above) containing 0.1% bovine serum albumin (BSA). Excess solution was carefully removed and slides were incubated with 1 nM [ $^{18}$ F]-PSS223 alone or together with 100 nM ABP688 or 100 nM JNJ16259685 in HEPES buffer for 45 min at ambient temperature. After incubation, the solutions were decanted and the slides washed on ice in HEPES buffer containing 0.1% BSA, and twice in HEPES buffer (3 minutes each) and finally dipped in H<sub>2</sub>O. Dried slides were exposed to a phosphor imager plate for 30 min and the plate was scanned in a BAS5000 reader (Fuji).

#### *Stability in PBS and plasma*

[ $^{18}$ F]-PSS223 (7-8 MBq) was incubated in phosphate buffer (4 mM KH<sub>2</sub>PO<sub>4</sub>/Na<sub>2</sub>HPO<sub>4</sub>, 155 mM NaCl, pH 7.4) or rat plasma at 37 °C for up to 2 h. At different time points, samples were diluted and enzymatic reactions stopped with ice cold MeCN (140 mL). Plasma samples were centrifuged at 12000xg for 10 min. The samples were filtered and supernatants were analyzed by UPLC.

#### *Microsome stability assay*

[ $^{18}$ F]-PSS223 (15 MBq) was preincubated for 5 min at 37 °C with 50 mL NADPH regenerating system A, 10 mL NADPH regenerating system B,

200 mL phosphate buffer (0.5 M, pH 7.4) and H<sub>2</sub>O in a total volume of 975 mL. Rat or human liver microsomes (25 mL) were added at a final protein concentration of 0.5 mg/mL at time 0. The mixture was incubated at 37 °C at 600 rpm and 100 mL aliquots were removed at the indicated time points and immediately quenched with the same volume of ice-cold MeCN. Proteins were precipitated at 12000xg (3 min) and the supernatants were filtered and analyzed by UPLC. NADPH regenerating system and microsomes were replaced by water, respectively in two control experiments.

#### *PET scans*

Rats were immobilized by anaesthesia with 2-3% isoflurane in oxygen/air on a GE Vista explore PET/CT scanner with the head in the field of view. Body temperature was controlled with a rectal probe connected to a 37 °C air blower and respiratory frequency was monitored with a 1025T Small Animal Monitoring and Gating System from SA Instruments (Sony Brook, NY, USA). At the start of data acquisition 15-30 MBq [ $^{18}$ F]-PSS223 or [ $^{11}$ C]-ABP688 was injected into a tail vein and data were collected in list mode for 90 and 60 min, respectively. After the PET scan, a CT was performed for anatomical orientation. PET data were reconstructed with 2D ordered subset expectation maximization (2D OSEM) and analyzed with PMOD 3.2 (PMOD, Zurich, Switzerland).

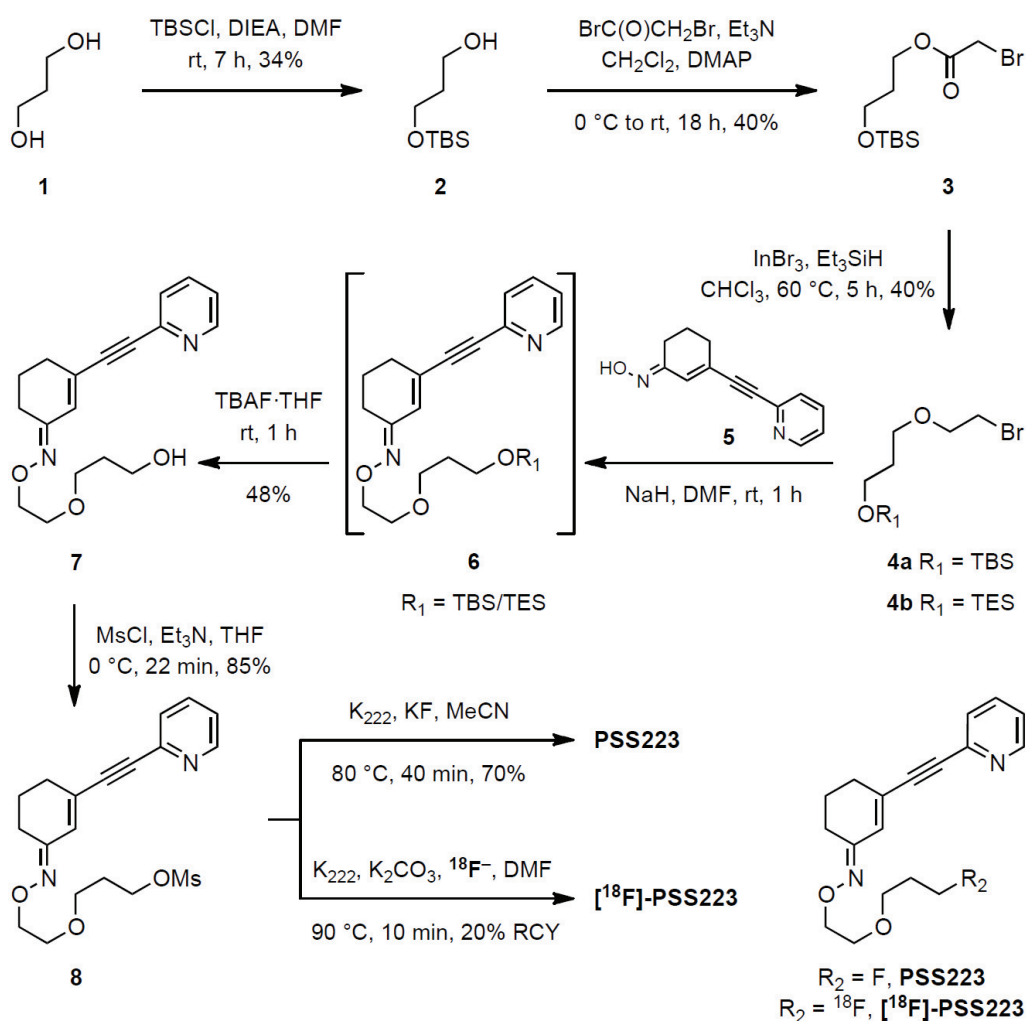
## **Results**

#### *Synthesis and radiolabelling*

The syntheses of the cold reference compound PSS223 and the desired radiolabelled analogue were accomplished via key intermediate **8** which was envisioned to originate from oxime **5** via S<sub>N</sub>2 reaction with bromoether **4**. Oxime **5** was prepared by a Sonogashira reaction [36] using commercially available 2-bromopyridine and (*E*)-3-ethynylcyclohex-2-enone oxime as previously reported [33] ([Supporting data](#)).

Monoprotection of commercially available propenediol with TBSCl, followed by esterification of the free hydroxyl group afforded ester **3** in 40% yield. Ester **3** was then reduced to the corresponding ether with triethylsilane [37] in a reaction catalyzed by indium(III)-bromide (**Figure 2**).

## Development of novel mGluR5 PET tracer



**Figure 2.** Syntheses of PSS223 and [ $^{18}\text{F}$ ]-PSS223 from key intermediate **8** via  $\text{S}_{\text{N}}2$  reaction.

Bromoethers **4a** and **4b** were reacted with oxime **5** to afford alcohol **7** after deprotection of silyl ethers with TBAF in 48% yield. Finally, mesylation of **7** afforded the desired precursor **8** which was used for the syntheses of PSS223 and [ $^{18}\text{F}$ ]-PSS223 (**Figure 2**). In the presence of Kryptofix-222<sup>®</sup>, nucleophilic substitution of mesylate **8** with potassium fluoride gave reference compound PSS223 in 70% yield. Under analogous conditions [ $^{18}\text{F}$ ]-PSS223 was obtained in 20% radiochemical decay corrected yield. The identity of [ $^{18}\text{F}$ ]-PSS223 was confirmed by HPLC analysis *via* co-injection with cold PSS223.

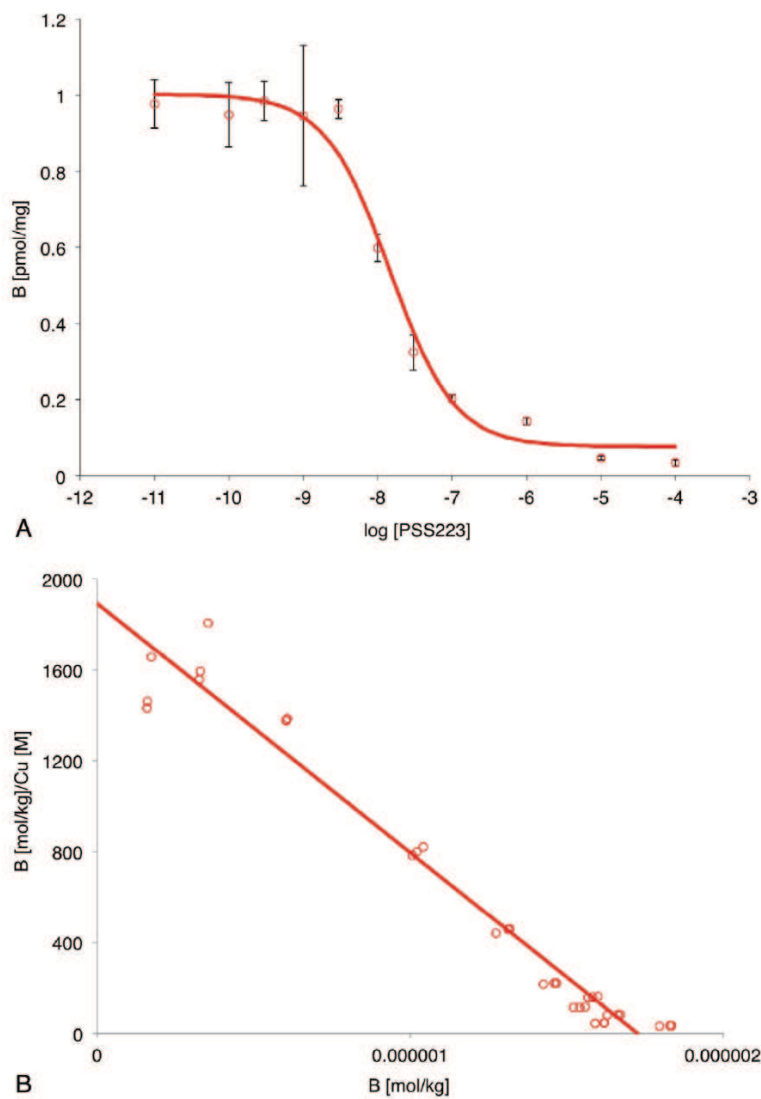
### *In vitro* evaluation

The experimentally determined  $\log D_{\text{pH}7.4}$  of [ $^{18}\text{F}$ ]-PSS223 was  $1.89 \pm 0.05$ . The binding affinity of

PSS223 was first estimated from a single competition binding experiment with 2 nM [ $^3\text{H}$ ]-ABP688 (**Figure 3A**). The estimated  $\text{IC}_{50}$  value was 14 nM and the respective  $K_i$  value amounted to 6 nM. The maximal displacement of [ $^3\text{H}$ ]-ABP688 by PSS223 was similar as observed with 100  $\mu\text{M}$  MMPEP, i.e., about 96% of total binding. Next, we determined  $K_d$  of [ $^{18}\text{F}$ ]-PSS223 in a saturation binding assay. Non-linear curve fitting of three independent experiments revealed a  $K_d$  of  $3.34 \pm 2.05$  nM and  $B_{\text{max}}$  ranging from 2 to 6 pmol/mg. Linearization in the Scatchard plot confirmed a single binding site (**Figure 3B**). Non-specific binding was estimated with an excess of MMPEP [38, 39].

*In vitro* autoradiography with rat brain slices showed heterogeneous radioactivity distribution





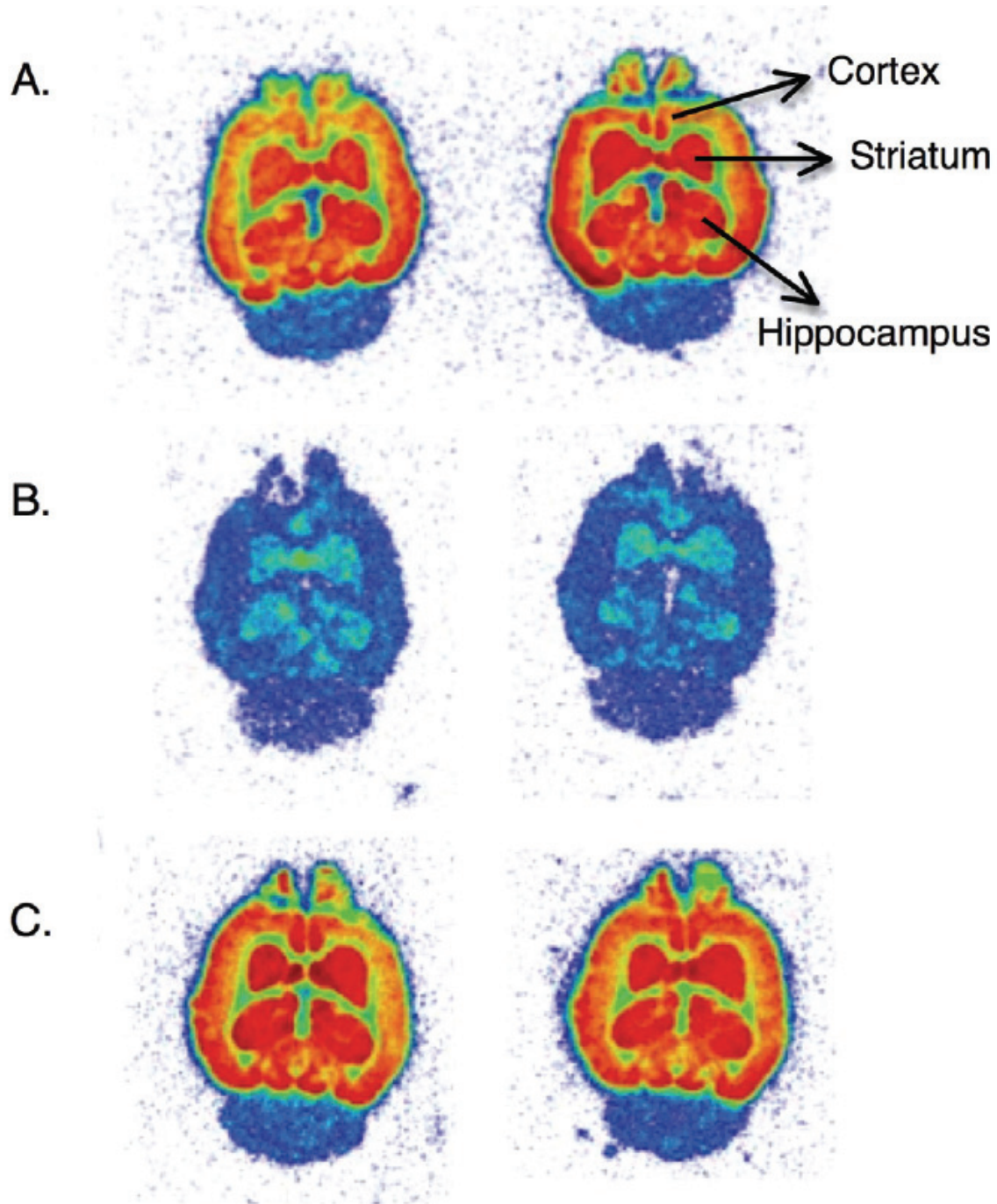
**Figure 3.** A. Displacement of  $[^3\text{H}]\text{-ABP688}$  by PSS223.  $[^3\text{H}]\text{-ABP688}$  (total 2 nM) binding (B) as a function of the PSS223 concentration. Symbols represent mean $\pm$ SD of one experiment performed in triplicate. Non-linear fitting (red line) resulted in  $\text{IC}_{50}$  14 nM and corresponding  $K_i$  6 nM; B. Scatchard plot analysis of the specific binding of  $[^{18}\text{F}]\text{-PSS223}$  to rat brain membranes. Symbols, specific binding (total binding minus binding in the presence of 100  $\mu\text{M}$  MMPEP) of one representative experiment; red line, linear regression.

after incubation with 1 nM  $[^{18}\text{F}]\text{-PSS223}$  (**Figure 4A**). Highest radioactivity was observed in striatum, hippocampus and cortex, regions known to express high levels of mGluR5 [40-42]. As expected from low mGluR5 expression in cerebellum [5], binding to this brain region was negligible. **Figure 4B** shows the displacement of  $[^{18}\text{F}]\text{-PSS223}$  with an excess of ABP688, confirming the specificity of  $[^{18}\text{F}]\text{-PSS223}$  for mGluR5. To demonstrate selectivity of  $[^{18}\text{F}]\text{-PSS223}$  for mGluR5 over mGluR1, the receptor with closest

sequence similarity to mGluR5, brain slices were incubated with 1 nM  $[^{18}\text{F}]\text{-PSS223}$  and an excess of the mGluR1 antagonist JNJ16259685 (**Figure 4C**). Binding competition was not observed in this case, confirming the selectivity of  $[^{18}\text{F}]\text{-PSS223}$  for mGluR5.

$[^{18}\text{F}]\text{-PSS223}$  was stable over 120 minutes in phosphate buffer and rat plasma at 37 °C. However, incubation with rat liver microsomes and NADPH under aerobic conditions resulted in

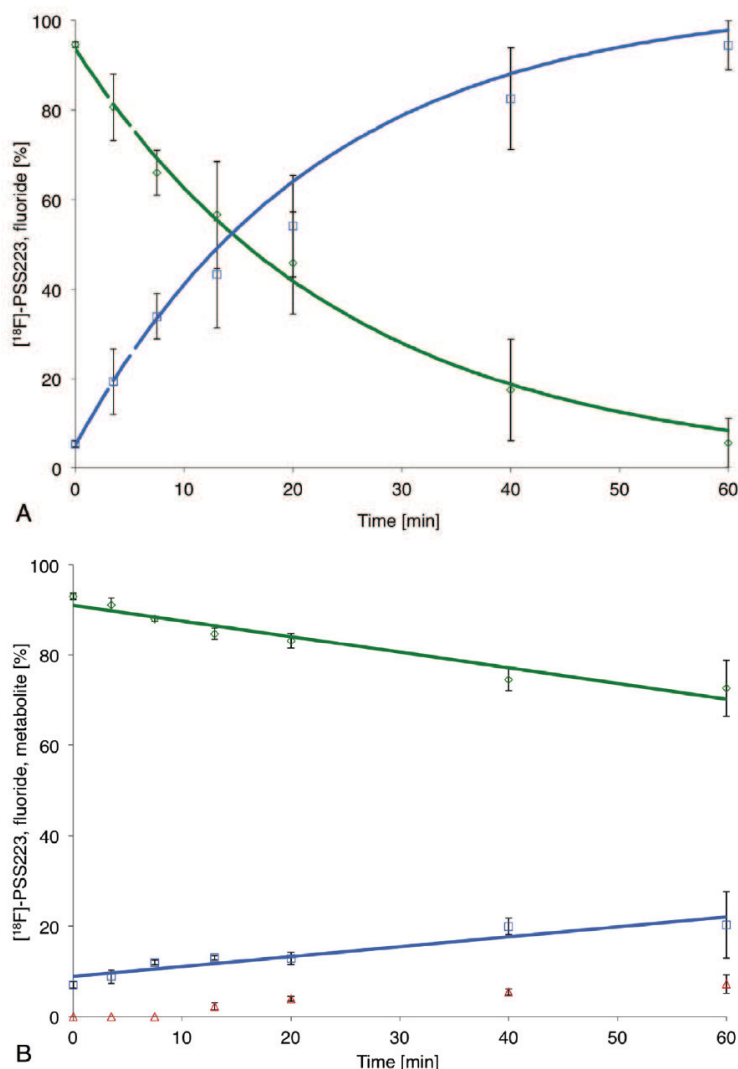




**Figure 4.** In vitro autoradiography with rat brain slices. A. Incubation with 1 nM [ $^{18}\text{F}$ ]-PSS223 showing heterogeneous distribution with highest expression in striatum, hippocampus and cortex; B. Brain slices incubated with 1 nM [ $^{18}\text{F}$ ]-PSS223 and 100 nM ABP688; C. Brain slices incubated with 1 nM [ $^{18}\text{F}$ ]-PSS223 and 100 nM mGluR1 antagonist JNJ16259685. Blue, green, orange, red, dark red indicate radioactivity from low to high. Columns represent same experiment performed in duplicate.

a significant decrease of [ $^{18}\text{F}$ ]-PSS223 and the appearance of a polar radiometabolite, co-

eluting with co-injected [ $^{18}\text{F}$ ]-fluoride. After one hour, about 90% of [ $^{18}\text{F}$ ]-PSS223 was metabo-



**Figure 5.** A. Metabolism of  $[^{18}\text{F}]$ -PSS223 by rat liver microsomes. Green symbols,  $[^{18}\text{F}]$ -PSS223; blue symbols, polar  $^{18}\text{F}$ -radiometabolite; lines, fitted exponential functions; B. Metabolism of  $[^{18}\text{F}]$ -PSS223 by human liver microsomes. Green symbols,  $[^{18}\text{F}]$ -PSS223; blue and red symbols, polar  $^{18}\text{F}$ -radiometabolites.

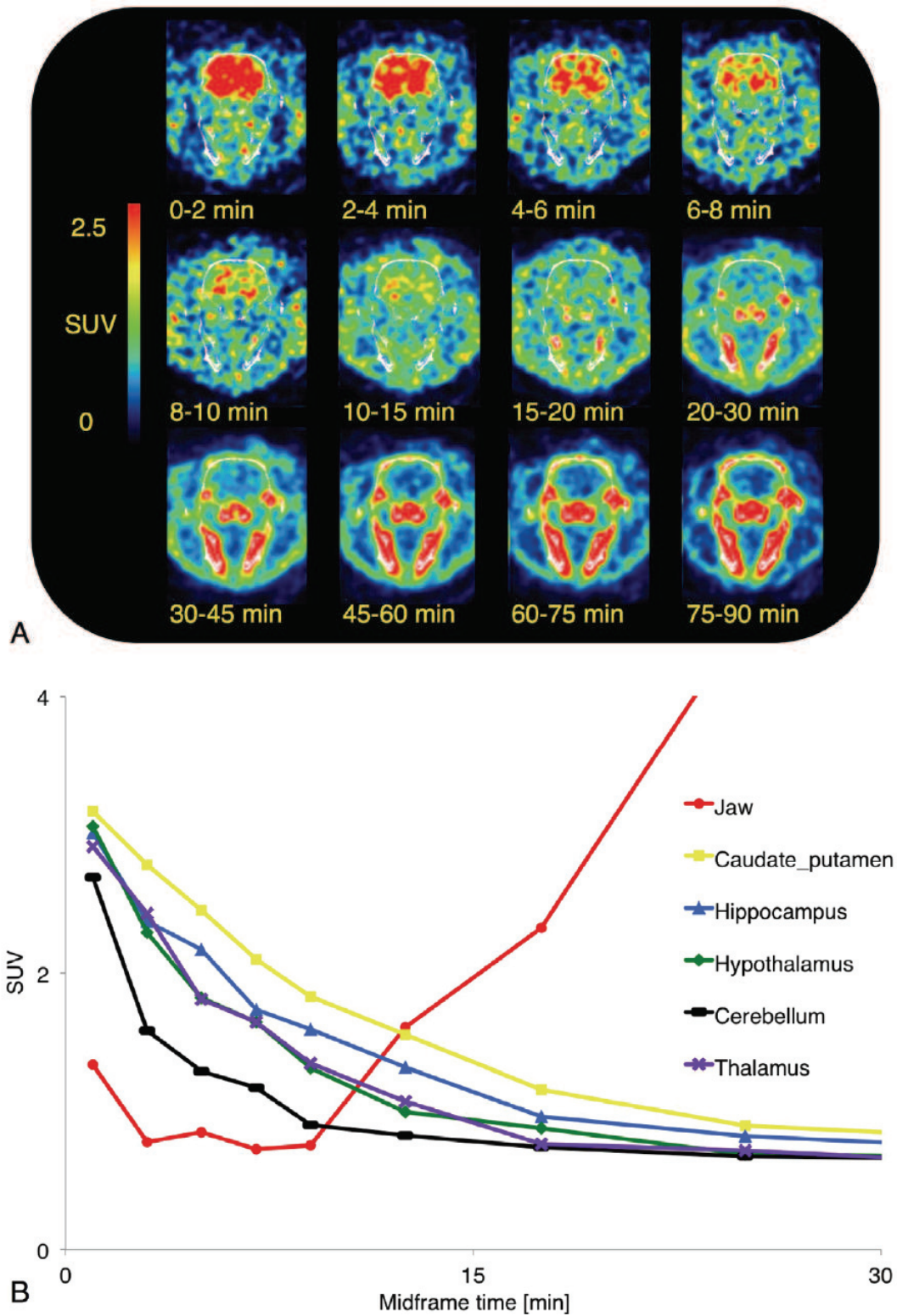
lized under the chosen experimental conditions (**Figure 5A**). Incubation of  $[^{18}\text{F}]$ -PSS223 with human liver microsomes under otherwise analogous conditions resulted in 20% decrease of  $[^{18}\text{F}]$ -PSS223 after one hour (**Figure 5B**) and the generation of two polar radiometabolites, the more polar one co-eluting with  $[^{18}\text{F}]$ -fluoride. None of the polar compounds were detected in the absence of microsomes or NADPH.

#### *In vivo evaluation*

Dynamic PET scans were performed with two rats to determine whether  $[^{18}\text{F}]$ -PSS223 could be used to image brain regions expressing mGluR5. **Figure 6A** shows a transversal plane across the head region over time. In the early time frames, brain perfusion resulted in high homogenous radioactivity in the brain. This was followed by a rapid washout from all brain regions and radioactivity accumulation in the skull and jaws. The time activity curves of mGluR5-rich brain regions and cerebellum show accumulation of  $[^{18}\text{F}]$ -PSS223 in striatum, hippocampus and cortex (**Figure 6B**). However, the mean residence time was relatively short and radioactivity approached cerebellum levels already after about 20 minutes. High accumulation of  $^{18}\text{F}$ -radioactivity in bone is demonstrated by the time activity curve of the jaws.

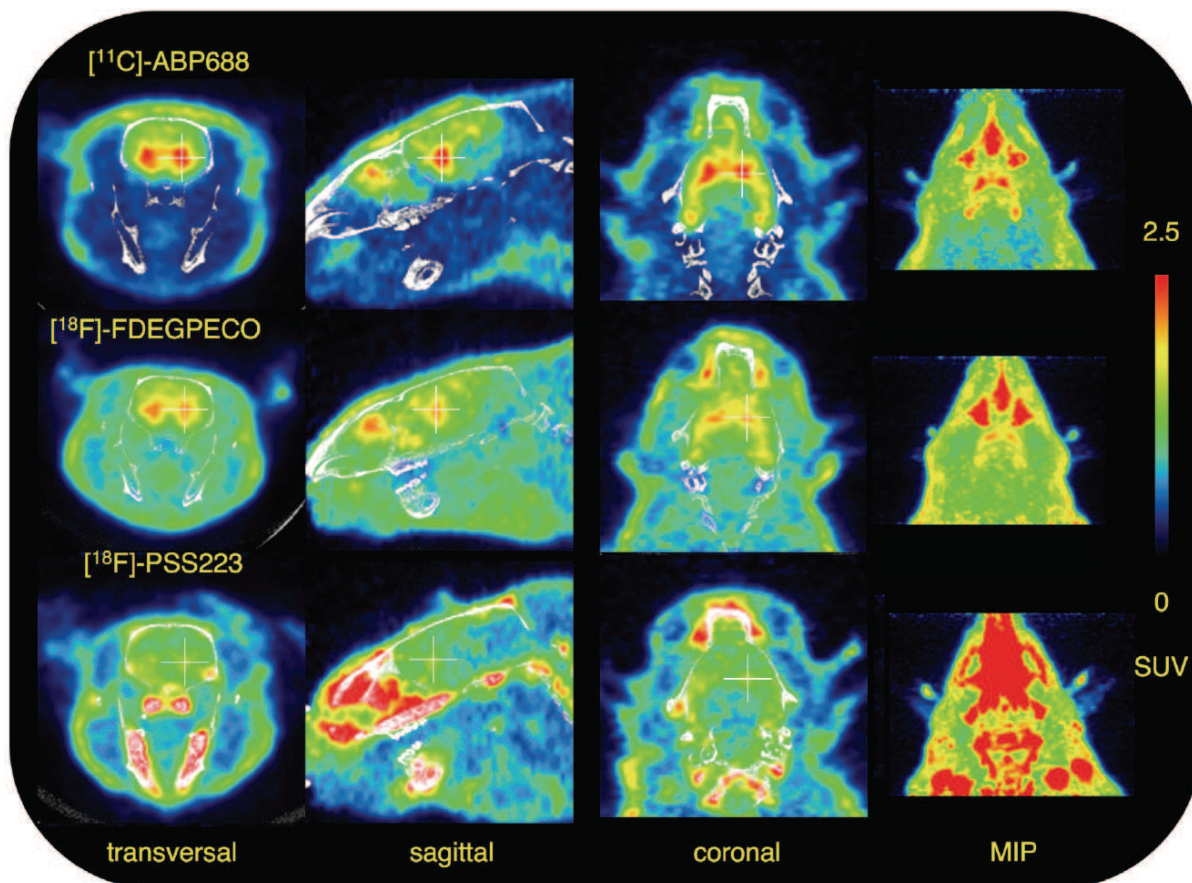
**Figure 7** shows a comparison of PET scans with  $[^{11}\text{C}]$ -ABP688,  $[^{18}\text{F}]$ -FDEGPeco and  $[^{18}\text{F}]$ -PSS223, all averaged from 2 to 45 min after tracer injection.  $[^{18}\text{F}]$ -PSS223 data are from the same scan shown in **Figures 6A** and B. The  $[^{18}\text{F}]$ -FDEGPeco image was reconstructed from the data shown in our recent publica-

tion [29, 43]. Comparing the three tracers,  $[^{11}\text{C}]$ -ABP688 showed the highest relative radioactivity accumulation in mGluR5-rich brain regions such as hippocampus and striatum.  $[^{18}\text{F}]$ -FDEGPeco also allows visualization of mGluR5-rich regions in the brain, however, at a higher background radioactivity. Our newest derivative,  $[^{18}\text{F}]$ -PSS223 shows only weak accumulation in mGluR5-rich brain regions and significant accu-



**Figure 6.** A. PET dynamic scan of the head of a male Wistar rat injected with  $[^{18}\text{F}]\text{-PSS223}$ . Transverse planes through the head summarized for the indicated time windows. White, CT; B. Time activity curves of  $[^{18}\text{F}]\text{-PSS223}$  uptake in a male Wistar rat in different brain regions showing increasing uptake in jaw due to rapid defluorination.





**Figure 7.** Rat brain PET images of [ $^{11}\text{C}$ ]-ABP688, [ $^{18}\text{F}$ ]-FDEGPeco and [ $^{18}\text{F}$ ]-PSS223. Planes (crosshairs) and maximal intensity projections (MIP) averaged from 2 to 45 min after tracer injection.

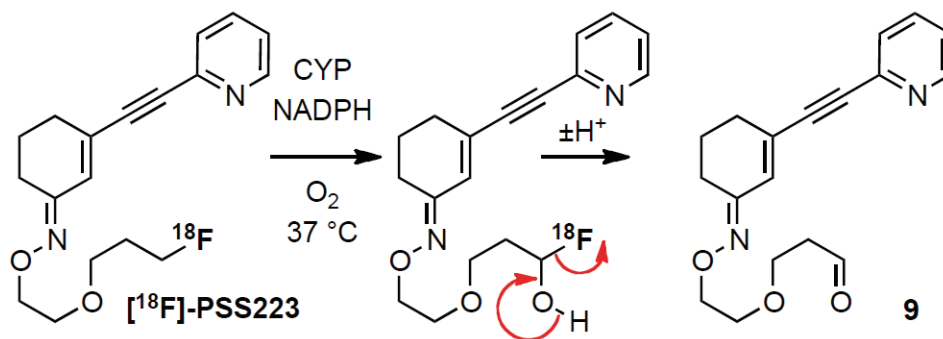
mulation of radioactivity in the skull and jaws as a result of rapid defluorination.

### Discussion

Reference compound PSS223 was prepared in an overall satisfactory yield and all compounds *en route* were fully characterized (NMR, IR, MS). The synthesis of bromoether **4** proved to be more challenging than initially anticipated. After several unsuccessful attempts to obtain compound **4** via direct functionalization with dihaloethane, an alternative approach was employed and yielded the desired bromoether **4** in 5% overall yield from three steps. During the reduction of the ester functionality in **3**, the TBS silyl ether was partially substituted by the TES group (originating from the reagent triethylsilane) to afford a mixture of TBS:TES silyl ethers in a ratio of 2:1 (**4a**:**4b**, Figure 2). For our pur-

poses, it was not necessary to separate the silyl ethers **4a** and **4b** since they were removed to afford free hydroxyl group in the following step.

The radiosynthesis of [ $^{18}\text{F}$ ]-PSS223 was accomplished in a single step from mesylate precursor **8** reproducibly in a good radiochemical yield. Pure radiotracer was obtained after HPLC semi-preparative purification in a total radiosynthesis time of 90 min (from the end of bombardment). Experimentally determined logD value of [ $^{18}\text{F}$ ]-PSS223 was 0.2 log units higher than that of [ $^{18}\text{F}$ ]-FDEGPeco ( $1.7 \pm 0.1$ ) [28] and 0.5 log units lower than that of [ $^{11}\text{C}$ ]-ABP688 [18]. This was within the optimal lipophilicity range of high affinity ABP688 analogues, (i.e., ClogP value below 2.5 and logP between 0.9 and 2.5, range of values desired for fast blood-brain barrier passage) [32]. One may assume that for those ABP688 analogues logD<sub>pH7.4</sub> corresponds to



**Figure 8.** Likely mechanism of defluorination of [ $^{18}\text{F}$ ]-PSS223 involves cytochrome P450-catalyzed C-oxygenation. The aldehyde **9** is oxidized or reduced to the respective carboxylic acid or alcohol while [ $^{18}\text{F}$ ]-fluoride is accumulated in bone.

logP as both the pyridine and the oxime nitrogen atoms are non-protonated at pH 7.4.

[ $^{18}\text{F}$ ]-PSS223 exhibited similarly high affinity for the mGluR5 as [ $^{11}\text{C}$ ]-ABP688, i.e., 3.3 nM compared to 1.7 nM [18], suggesting that the long chain in [ $^{18}\text{F}$ ]-PSS223 did not significantly affect the binding affinity. The value of  $B_{\text{max}}$  determined for [ $^{18}\text{F}$ ]-PSS223 (range: 2-6 pmol/mg) compared favourably to the  $B_{\text{max}}$  values of [ $^{11}\text{C}$ ]-ABP688 ( $231 \pm 18$  fmol/mg) [18] and [ $^{18}\text{F}$ ]-FDEGPECO (range: 0.5-4 pmol/mg) [28]. Specific binding of [ $^{18}\text{F}$ ]-PSS223 to mGluR5-rich brain regions in the *in vitro* autoradiography experiments prompted further *in vitro* and *in vivo* characterization of [ $^{18}\text{F}$ ]-PSS223.

[ $^{18}\text{F}$ ]-PSS223 was stable in buffer and rat plasma; however, it was significantly metabolized by the rat liver microsomal enzymes and a polar radiometabolite co-eluting with [ $^{18}\text{F}$ ]-fluoride was observed by UPLC. In the experiments with human liver microsomes, two polar radiometabolites were detected albeit in the amounts significantly lower than those observed with rat microsomal enzymes. Based on the control experiments, it was concluded that the process was NADPH-dependent, implying the involvement of oxidoreductases. One likely mechanism involves defluorination preceded by the oxygenation of the carbon atom in the  $\alpha$ -position to the fluorine atom [44]. The possible mechanism is depicted in **Figure 8**. A lower degree of metabolic activity of the human compared to that of rat liver microsomes would be in agreement with several other studies [45-48].

Considering the rapid metabolism *in vitro* and the possibility of defluorination *in vivo*, we performed two dynamic PET scans. PET analysis demonstrated rapid wash-out of radioactivity from the brain and high accumulation in the skull and jaws. Accumulation of [ $^{18}\text{F}$ ]-PSS223 in mGluR5-rich brain regions was significantly lower than observed for [ $^{18}\text{F}$ ]-FDEGPECO. [ $^{18}\text{F}$ ]-PSS223 was therefore, not further investigated *in vivo*. The observed radioactivity accumulation in bone supports the conclusion that the radiometabolite (or one of the radiometabolites) observed with liver microsomes was [ $^{18}\text{F}$ ]-fluoride. Based on the higher stability of [ $^{18}\text{F}$ ]-PSS223 in human liver microsomes, less extensive defluorination is expected in humans.

The difference in the *in vivo* behaviour between [ $^{18}\text{F}$ ]-PSS223 and [ $^{18}\text{F}$ ]-FDEGPECO could be attributed to the  $\beta$ -heteroatom effect [49, 50], by which primary aliphatic  $^{18}\text{F}$ -atoms in a  $\beta$ -position to heteroatom (e.g., [ $^{18}\text{F}$ ]-FCH<sub>2</sub>CH<sub>2</sub>OR) are found to be metabolized at a slower rate. This rationale supports absence of defluorination for [ $^{18}\text{F}$ ]-FDEGPECO.

## Conclusions

In conclusion, the radiosynthesis of [ $^{18}\text{F}$ ]-PSS223 was accomplished in good radiochemical yields and high specific radioactivity. The new radioligand binds mGluR5 *in vitro* with low nanomolar affinity and shows heterogeneous and specific accumulation *in vitro* in mGluR5-rich brain regions in autoradiographic studies. Although the PET studies with [ $^{18}\text{F}$ ]-PSS223 did

not allow a clear-cut visualization of mGluR5 *in vivo* in the rat brain, due to low metabolic stability and rapid wash-out, [<sup>18</sup>F]-PSS223 could potentially find utility in higher animals considering that [<sup>18</sup>F]-PSS223 exhibited higher stability in human microsomes.

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Experimental procedures and characterization data for all compounds, and HPLC chromatographs of [<sup>18</sup>F]-PSS223 are provided.

## Disclaimer of conflict of interest

none.

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# Synthesis, Radiolabelling and *In Vitro* and *In Vivo* Evaluation of a Novel Fluorinated ABP688 Derivative for the PET Imaging of Metabotropic Glutamate Receptor Subtype 5

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## Supporting Information

### Chemistry

**3-((*Tert*-butyldimethylsilyl)oxy)propan-1-ol (2):** A flame dried flask was charged with anhydrous *N,N*'-dimethylformamide (25 mL) and at ambient temperature under N<sub>2</sub> atmosphere 1,3-propanediol (1.00 mL, 1.05 g, 13.8 mmol, d=1.05) was added and the resulting colourless solution was treated with diisopropylethylamine (22.5 mL, 17.0 g, 131 mmol, d=0.755) in one portion and pale yellow biphasic mixture was vigorously stirred and further treated with a solution of *tert*-butyldimethylchlorosilane (2.08 g, 13.8 mmol) in DMF (15 mL) dropwise over 16 min during which time the mixture turned cloudy and it was allowed to stir for 6 h. After this time the mixture was partitioned between H<sub>2</sub>O (30 mL) and Et<sub>2</sub>O (50 mL) and the two layers were well shaken and separated. The aqueous phase was extracted with Et<sub>2</sub>O (2x50 mL). The combined organic extracts were washed with 2M aq. HCl (2x50 mL; CAUTION: vigorous bubbling), saturated aq. NaHCO<sub>3</sub> (1x50 mL), brine (1x50 mL), dried (Na<sub>2</sub>SO<sub>4</sub>) and concentrated *in vacuo* to give crude mixture as pale yellow oil (3.78 g). The crude product was purified by chromatography on a silica gel column (eluting with EtOAc:pentane 3:7) to afford title compound (1.29 g, 6.77 mmol, 49%) as a colourless oil: IR (neat) 3358, 2954, 2930, 1741, 1472, 1373, 1251, 1094, 962, 835, 776 cm<sup>-1</sup>; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 3.84 (t, *J* = 5.6 Hz, 2H), 3.80 (dd, *J* = 10.9, 5.4 Hz, 2H), 2.54 (t, *J* = 5.3 Hz, 1H), 1.78 (quint, *J* = 5.6 Hz, 1H), 0.90 (s, 9H), 0.08 (s, 6H) ppm; <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ 63.0 (2), 62.6 (2), 34.3 (2), 26.1 (3C, 3), 18.4 (0), -5.3 (2C, 3) ppm; MS (ES<sup>+</sup>) *m/z* 191 (M + H)<sup>+</sup>; HRMS (EI<sup>+</sup>) *m/z* 191.1462 (calcd. for C<sub>9</sub>H<sub>23</sub>O<sub>2</sub>Si: 191.1467).

**3-((*Tert*-butyldimethylsilyl)oxy)propyl 2-bromoacetate (3):** A stirred solution of 3-((*tert*-butyldimethylsilyl)oxy)propan-1-ol (818 mg, 4.30 mmol) in anhydrous dichloromethane (9 mL) was allowed to cool to 0 °C (the ice bath) and under N<sub>2</sub> atmosphere it was then treated with triethylamine (0.66 mL, 478 mg, 4.73 mmol, d=0.726) in one portion followed by 4-dimethylaminopyridine (26.0 mg, 0.22 mmol) and finally bromoacetyl bromide (0.38 mL, 867 mg, 4.30 mmol, d=2.31) was added dropwise over 10 min during which time the mixture turned orange and then yellow and cloudy and it was allowed to stir and slowly warm to ambient temperature over 16 h. After this time the mixture was partitioned between H<sub>2</sub>O (20 mL) and CH<sub>2</sub>Cl<sub>2</sub> (10 mL) and the two layers were well shaken and separated. The aqueous phase was extracted with CH<sub>2</sub>Cl<sub>2</sub> (2x20 mL). The combined organic extracts were washed with H<sub>2</sub>O (3x20 mL), brine (1x20 mL), dried (Na<sub>2</sub>SO<sub>4</sub>) and concentrated *in vacuo* to give crude mixture as brown oily residue (2.49 g). The crude mixture was purified by

chromatography on a silica gel column (eluting with gradient EtOAc:pentane 1:18 to EtOAc:pentane 1:9) to afford title compound (489 mg, 1.57 mmol, 36%) as a colourless oil: IR (neat) 2954, 2931, 2854, 1742, 1473, 1277, 1257, 1103, 1008, 969, 834  $\text{cm}^{-1}$ ;  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$  4.28 (t,  $J$  = 6.4 Hz, 2H), 3.83 (s, 2H), 3.71 (t,  $J$  = 6.0 Hz, 2H), 1.87 (quint,  $J$  = 6.2 Hz, 1H), 0.89 (s, 9H), 0.05 (s, 6H) ppm;  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ )  $\delta$  167.4 (O), 63.5 (2), 59.3 (2), 31.7 (2), 26.1 (3C, 3), 26.0 (2), 18.5 (O), -5.2 (2C, 3) ppm; HRMS (EI+)  $m/z$  252.9890 ((M -  $\text{C}_4\text{H}_9$ ) $^+$  calcd. for  $\text{C}_7\text{H}_{14}\text{BrO}_3\text{Si}$ ; 252.9896).

**(3-(2-Bromoethoxy)propoxy)(tert-butyl)dimethylsilane (4a):** A stirred solution of 3-((tert-butyl)dimethylsilyl)oxy)propyl 2-bromoacetate (709 mg, 2.28 mmol) in chloroform (2.2 mL) under  $\text{N}_2$  atmosphere was treated with indium(III) bromide (40.4 mg, 0.11 mmol) in one portion and triethylsilane (1.46 mL, 1.06 g, 9.12 mmol,  $d=0.728$ ) was then added dropwise (addition time < 1 min) and the resulting heterogeneous mixture was allowed to heat at 60  $^\circ\text{C}$  (oil bath temperature) over 5 h in a flask equipped with condenser. Immediately upon heating mixture turned creamy and yellow. After this time the mixture was allowed to cool to ambient temperature and it was then diluted with  $\text{H}_2\text{O}$  (12 mL) and  $\text{CH}_2\text{Cl}_2$  (20 mL) and the two layers were well shaken and separated. The aqueous phase was further extracted with  $\text{CH}_2\text{Cl}_2$  (2x20 mL). The combined organic extracts were washed with brine (1x30 mL), dried ( $\text{Na}_2\text{SO}_4$ ) and concentrated *in vacuo* to give crude mixture as brown oily residue (2.49 g). The crude product was purified by chromatography on a silica gel column (eluting with EtOAc:pentane 1:18) to afford the title compound (287 mg, 0.97 mmol, 42%) as a mixture with **(3-(2-bromoethoxy)propoxy)triethylsilane (4b)** in a 5:1 NMR ratio, respectively: IR (neat) 2954, 2928, 2857, 1471, 1255, 1098, 1006, 836, 776, 745, 666  $\text{cm}^{-1}$ ;  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$  3.74 (t,  $J$  = 6.3 Hz, 4H, overlapped), 3.71 (t,  $J$  = 6.2 Hz, 4H, overlapped), 3.59 (t,  $J$  = 6.2 Hz, 2H, **4b**), 3.58 (t,  $J$  = 6.3 Hz, 2H, **4a**), 3.46 (t,  $J$  = 6.3 Hz, 4H, overlapped), 1.81 (quint,  $J$  = 6.2 Hz, 4H, **4b**), 1.79 (quint,  $J$  = 6.2 Hz, 4H, **4a**), 0.96 (t,  $J$  = 8.0 Hz, 9H, **4b**), 0.89 (s, 9H, **4a**), 0.60 (q,  $J$  = 8.0 Hz, 6H, **4b**), 0.05 (s, 6H, **4a**) ppm; MS (EI+)  $m/z$  239 (M -  $\text{C}_4\text{H}_9$ ) $^+$  for **4a** and 267 (M -  $\text{C}_2\text{H}_5$ ) $^+$  for **4b**. This mixture was used for the next step.

**(E)-3-(Pyridin-2-ylethynyl)cyclohex-2-enone O-(2-(3-((tert-butyl)dimethylsilyl)oxy)propoxy)ethyl) oxime (6a) and (E)-3-(pyridin-2-ylethynyl)cyclohex-2-enone O-(2-(3-(triethylsilyl)oxy)propoxy)ethyl) oxime (6b):** A flame dried flask was charged with (E)-3-(pyridin-2-ylethynyl)cyclohex-2-enone oxime (185 mg, 0.87 mmol) and anhydrous  $N,N'$ -dimethylformamide (10 mL) was added and clear pale yellow mixture was treated with sodiumhydride (50 mg of 60% suspension in oil, 1.25 mmol) and the resulting bright yellow heterogeneous mixture was stirred at ambient temperature under  $\text{N}_2$  atmosphere for 36 min during which time the mixture turned orange. After this time a mixture of (3-(2-bromoethoxy)propoxy)(tert-butyl)dimethylsilane and (3-(2-bromoethoxy)propoxy)triethylsilane (5:1 ratio, respectively, 287 mg, 0.96 mmol) in DMF (7.5 mL) was added dropwise over 8 min during which time the mixture turned brown and it was allowed to stir further for 70 min. After this time the crude mixture was quenched with saturated aq.  $\text{NaHCO}_3$  (10 mL) and it was diluted with  $\text{H}_2\text{O}$  (40 mL) and  $\text{Et}_2\text{O}$  (80 mL) and the two layers were well shaken and separated. The aqueous phase was extracted with  $\text{Et}_2\text{O}$  (2x80 mL). The combined organic extracts were washed with  $\text{H}_2\text{O}$  (3x40 mL), brine (1x40 mL),

dried (Na<sub>2</sub>SO<sub>4</sub>) and concentrated *in vacuo* to give the crude mixture as a brown oily residue (293 mg). The crude product was used for the next step without purification.

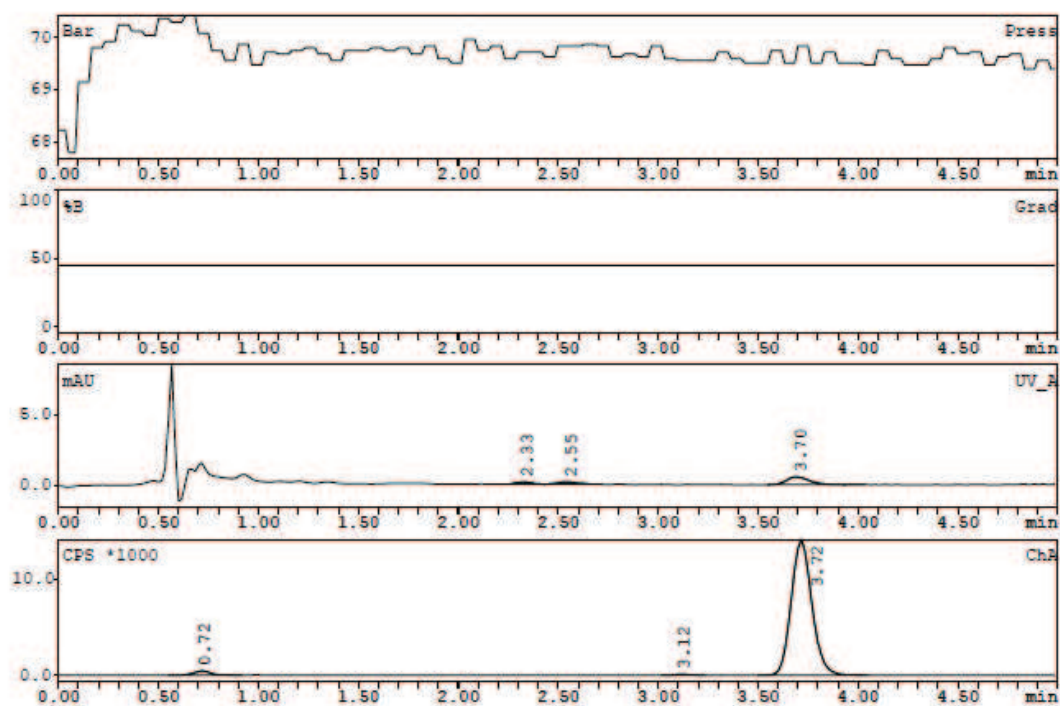
**(E)-3-(Pyridin-2-ylethynyl)cyclohex-2-enone O-(2-(3-hydroxypropoxy)ethyl) oxime (7):** At ambient temperature under N<sub>2</sub> atmosphere round bottom flask was charged with the crude mixture of (E)-3-(pyridin-2-ylethynyl)cyclohex-2-enone O-(2-(3-((tert-butyldimethylsilyl)oxy)propoxy)ethyl) oxime and (E)-3-(pyridin-2-ylethynyl)cyclohex-2-enone O-(2-(3-((triethylsilyl)oxy)propoxy)ethyl) oxime (293 mg, 0.68 mmol) and anhydrous tetrahydrofuran (12.3 mL) was added and the resulting clear orange mixture was further treated with tetrabutylammoniumfluoride solution in THF (1.4 mL, 1.36 mmol, c=1M) dropwise over 5 min and the resulting brown mixture was allowed to stir at ambient temperature under N<sub>2</sub> for 70 min. After this time the mixture was partitioned between H<sub>2</sub>O (30 mL) and EtOAc (40 mL) and the two layers were well shaken and separated. The aqueous phase was extracted with EtOAc (2x40 mL, slow separation of phases). The combined organic extracts were washed with H<sub>2</sub>O (3x30 mL), brine (1x40 mL), dried (Na<sub>2</sub>SO<sub>4</sub>) and concentrated *in vacuo* to give crude mixture as brown oil. The crude product was purified by chromatography on a silica gel column (eluting with gradient EtOAc:pentane 9:1 to 100% EtOAc) to give the title compound (131 mg, 0.42 mmol) as a pale yellow oil: IR (neat) 3412, 2936, 2869, 2196, 1581, 1463, 1428, 1358, 1248, 1122, 1059, 978, 958, 861, 779 cm<sup>-1</sup>; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 8.60 (ddd, *J* = 4.9, 1.7, 0.9 Hz, 1H), 7.66 (td, *J* = 7.8, 1.8 Hz, 1H), 7.45 (dt, *J* = 7.8, 1.0 Hz, 1H), 7.23 (ddd, *J* = 7.6, 4.9, 1.2 Hz, 1H), 6.58 (t, *J* = 1.6 Hz, 1H), 4.26 (bdm, *J* = 4.6 Hz, 2H), 3.78 (t, *J* = 5.5 Hz, 2H), 3.71 (bdm, *J* = 4.8 Hz, 2H), 3.72 (bdm, *J* = 3.5 Hz, 2H), 3.69 (dd, *J* = 5.7 Hz, 2H), 2.57 (dd, *J* = 6.4 Hz, 2H), 2.40 (td, *J* = 6.1, 1.6 Hz, 2H), 1.84 (quint, *J* = 5.6 Hz, 2H), 1.80 (quint, *J* = 6.3 Hz, 2H) ppm; <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ 156.0 (O), 150.2 (1), 143.4 (O), 136.4 (1), 131.2 (1), 127.6 (O), 127.5 (1), 123.0 (1), 92.0 (O), 90.2 (O), 73.5 (2), 70.9 (2), 69.8 (2), 62.4 (2), 32.0 (2), 29.6 (2), 22.4 (2), 21.0 (2) ppm; MS (ES<sup>+</sup>) *m/z* 315 (M + H)<sup>+</sup>; HRMS (ESI) *m/z* 315.1710 (calcd. for C<sub>18</sub>H<sub>23</sub>N<sub>2</sub>O<sub>3</sub>: 315.1703).

**(E)-3-(2-(((3-(Pyridin-2-ylethynyl)cyclohex-2-en-1-ylidene)amino)oxy)ethoxy)propyl methanesulfonate (8):** A stirred solution of (E)-3-(pyridin-2-ylethynyl)cyclohex-2-enone O-(2-(3-hydroxypropoxy)ethyl) oxime (131 mg, 0.42 mmol) in anhydrous tetrahydrofuran (4.2 mL) was treated with triethylamine (39 µL, 57.0 mg, 0.50 mmol, d=1.477) in one portion and methanesulfonylchloride (117 µL, 85.0 mg, 0.84 mmol, d=0.726) was then added dropwise (addition time < 1 min) and the resulting pale yellow mixture was allowed to stir at ambient temperature under N<sub>2</sub> atmosphere for 22 min during which time white precipitate formed. After this time the mixture was partitioned between H<sub>2</sub>O (16 mL) and EtOAc (20 mL) and the two layers were well shaken and separated. The aqueous phase was extracted with EtOAc (2x20 mL). The combined organic extracts were washed with H<sub>2</sub>O (3x16 mL), brine (1x16 mL), dried (Na<sub>2</sub>SO<sub>4</sub>) and concentrated *in vacuo* to give crude mixture as pale yellow oil. The crude product was purified by chromatography on a silica gel column (eluting with gradient EtOAc:pentane 4:1 to 100% EtOAc) to afford the title compound (138 mg, 0.35 mmol, 85%) as a pale yellow oil: IR (neat) 2933, 2870, 2197, 2097, 1580, 1462, 1428, 1352, 1173, 1124, 1061, 887, 863, 842, 779, 629 cm<sup>-1</sup>; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 8.59 (bdm, *J* = 4.4 Hz, 1H), 7.65 (td, *J* = 7.8, 1.8 Hz, 1H), 7.44 (bdm, *J* = 7.8 Hz, 1H), 7.22 (ddd, *J* = 7.6, 4.9, 1.1 Hz, 1H), 6.56 (t, *J* = 1.5 Hz, 1H),

4.34 (t,  $J = 6.2$  Hz, 2H), 4.24 (bdm,  $J = 4.7$  Hz, 2H), 3.70 (bdm,  $J = 4.8$  Hz, 2H), 3.59 (t,  $J = 5.9$  Hz, 2H), 3.00 (s, 3H), 2.57 (dm,  $J = 6.5$  Hz, 2H), 2.40 (td,  $J = 6.2, 1.4$  Hz, 2H), 2.01 (quint,  $J = 6.0$  Hz, 2H), 1.80 (quint,  $J = 6.4$  Hz, 2H) ppm;  $^{13}\text{C}$  NMR (100MHz,  $\text{CDCl}_3$ )  $\delta$  155.9 (0), 150.3 (1), 143.4 (0), 136.3 (1), 131.1 (1), 127.6 (0), 127.4 (1), 123.0 (1), 92.1 (0), 90.0 (0), 73.7 (2), 69.6 (2), 67.5 (2), 66.6 (2), 37.3 (3), 29.6 (2), 29.5 (2), 22.5 (2), 20.9 (2) ppm; MS (ES+)  $m/z$  393 ( $\text{M} + \text{H}^+$ ); HRMS (ESI)  $m/z$  393.1489 (calcd. for  $\text{C}_{19}\text{H}_{25}\text{N}_2\text{O}_5\text{S}$ : 393.1479).

**(E)-3-(Pyridin-2-ylethynyl)cyclohex-2-enone O-(2-(3-fluoropropoxy)ethyl) oxime (PSS223):** A flame dried round bottom flask was charged with Kryptofix-222® (150 mg, 0.40 mmol) and potassium fluoride (23.0 mg, 0.40 mmol) and at ambient temperature under  $\text{N}_2$  atmosphere anhydrous acetonitrile (3.2 mL) was added. Resulting colourless solution was further treated with a solution of (E)-3-(2-(((3-(pyridin-2-ylethynyl)cyclohex-2-en-1-ylidene)amino)oxy)ethoxy)propyl methanesulfonate (78.0 mg, 0.20 mmol) in anhydrous acetonitrile (3.2 mL) dropwise over 2 min during which time the mixture turned pale orange and the mixture was allowed to heat at  $80^\circ\text{C}$  (oil bath temperature) for 40 min. The mixture was then allowed to cool to ambient temperature and then diluted with  $\text{H}_2\text{O}$  (15 mL) and EtOAc (25 mL) and the two layers were well shaken and separated. The aqueous phase was extracted with EtOAc (2x25 mL, slow separation of phases). The combined organic extracts were washed with  $\text{H}_2\text{O}$  (3x15 mL), brine (1x15 mL), dried ( $\text{Na}_2\text{SO}_4$ ) and concentrated *in vacuo* to give the title compound (44.1 mg, 0.14 mmol, 70%) as a colourless oil: IR (neat): 3050, 2925, 2869, 2205, 1580, 1462, 1358, 1124, 1060, 978, 864, 778, 741  $\text{cm}^{-1}$ ;  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$  8.59 (ddd,  $J = 4.9, 1.8, 0.9$  Hz, 1H), 7.65 (td,  $J = 7.8, 1.8$  Hz, 1H), 7.44 (dt,  $J = 7.8, 1.0$  Hz, 1H), 7.22 (ddd,  $J = 7.6, 5.0, 1.2$  Hz, 1H), 6.57 (t,  $J = 1.6$  Hz, 1H), 4.55 (dt,  $J = 4.7, 5.9$  Hz, 2H), 4.25 (dm,  $J = 4.8$  Hz, 2H), 3.71 (dm,  $J = 4.9$  Hz, 2H), 3.61 (t,  $J = 6.2$  Hz, 2H), 2.58 (ddm,  $J = 6.4$  Hz, 2H), 2.40 (td,  $J = 6.0, 1.5$  Hz, 2H), 1.96 (dq,  $J = 26, 6.0$  Hz, 2H), 1.80 (quint,  $J = 6.4$  Hz, 2H) ppm;  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ )  $\delta$  156.0 (0), 150.3 (1), 143.6 (0), 136.3 (1), 131.2 (1), 127.45 (0), 127.42 (1), 123.0 (1), 92.4 (0), 90.1 (0), 81.5 (d,  $J = 141$  Hz, 2), 73.8 (2), 69.6 (0), 67.0 (d,  $J = 5.5$  Hz, 2), 31.0 (d,  $J = 20$  Hz, 2), 29.6 (2), 22.5 (2), 21.0 (2) ppm;  $^{19}\text{F}$  NMR (376 MHz,  $\text{CDCl}_3$ )  $\delta$  -221.8 (ddt,  $J = 46, 25$  Hz) ppm; MS (ES+)  $m/z$  317 ( $\text{M} + \text{H}^+$ ); HRMS (ESI)  $m/z$  317.1663 (calcd. for  $\text{C}_{18}\text{H}_{22}\text{FN}_2\text{O}_2$ : 317.1660).

HPLC chromatograph of [ $^{18}\text{F}$ ]-PSS223: Quality control



## Sample description

Study: MGLUR5  
 Measurement: SDM-RS-40A, injection : 20.06.2011 10:38  
 Method: ISO45\_ACN IN 5MIN from: 01.04.2011 05:30 Position: 1  
 Station number: 2  
 Inj. volume (µl): 5.0  
 Radio detector: raytest Gabi Star Serial Nr.: #30168 raytest GINA star 20.04.09 V4.8

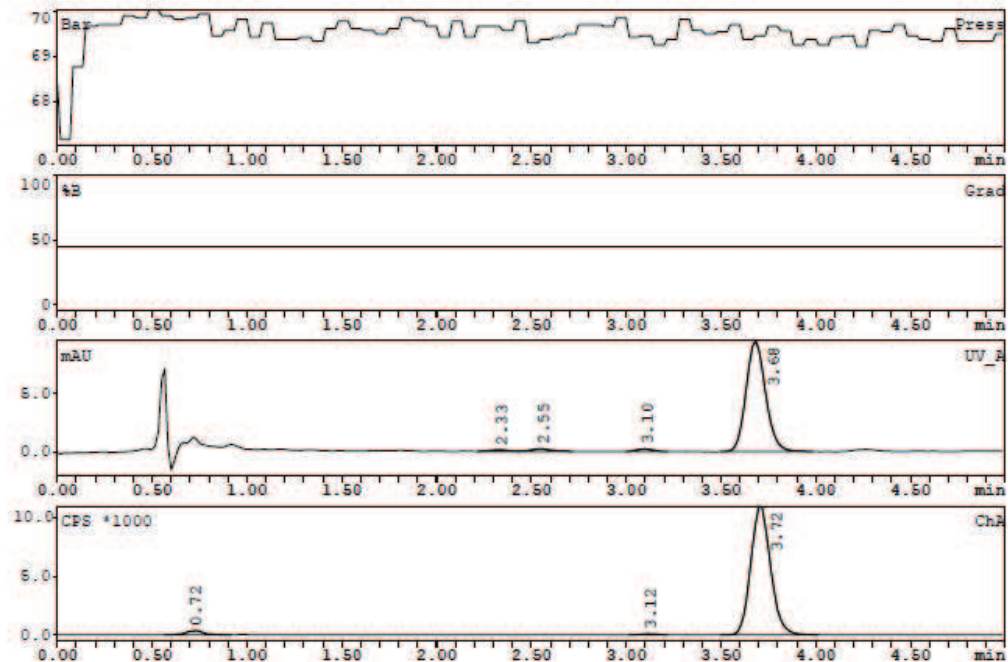
## Integration ChA

Substance	R/T min	Type	Area Counts	%Area %
Reg #1	0.72	BB(	2661.8	2.55
Reg #2	3.12	BB(	496.8	0.48
Reg #3	3.72	BB(	101096.1	96.97
Sum in ROI			104254.7	
Area			106831.4	
Ext. BKG			0.00 CPS	

## Integration UV\_A

Substance	R/T min	Type	Area mAU*min	%Area %
Reg #1	2.33	DB(	0.785731	12.57
Reg #2	2.55	DB(	1.003935	16.06
Reg #3	3.70	BB(	4.462353	71.37
Sum in ROI			6.252019	

HPLC chromatograph of [ $^{18}\text{F}$ ]-PSS223: Coinjection with cold reference



## Sample description

Study: MGLUR5  
 Measurement: SDM-RS-40CONIECTION, injection : 20.06.2011 10:49  
 Method: ISO45\_ACN IN 5MIN from: 01.04.2011 05:30: Position: 1  
 Station number: 2  
 Inj. volume (µl): 5.0  
 COINJECTION WITH COLD REFERENCE  
 Radio detector: raytest Gabi Star Serial Nr.: #30168 raytest GINA star 20.04.09 V4.8

## Integration ChA

Substance	R/T min	Type	Area Counts	%Area %
Reg #1	0.72	BB(	2254.93	2.73
Reg #2	3.12	BB(	410.08	0.50
Reg #3	3.72	BB(	79816.78	96.77
Sum in ROI			82483.79	
Area			85725.75	
Ext. BKG			0.00 CPS	

## Integration UV\_A

Substance	R/T min	Type	Area mAU*min	%Area %
Reg #1	2.33	BD(	0.82671	1.15
Reg #2	2.55	DB(	1.66488	2.33
Reg #3	3.10	BB(	1.28342	1.79
Reg #4	3.68	BB(	67.80675	94.73
Sum in ROI			71.58175	